

This article was downloaded by:

On: 15 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Chemistry and Ecology

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713455114>

A Functional Approach to the Assessment of the Nutritional Value of Particulate Organic Matter

Cristina Mistic^a; Mauro Fabiano^b

^a Istituto Scienze Ambientali Marine, Università di Genova, S. Margherita L., Italy ^b Cattedra di Ecologia, Università di Ancona, Ancona., Italy

To cite this Article Mistic, Cristina and Fabiano, Mauro(1996) 'A Functional Approach to the Assessment of the Nutritional Value of Particulate Organic Matter', *Chemistry and Ecology*, 13: 1, 51 – 63

To link to this Article: DOI: 10.1080/02757549608039101

URL: <http://dx.doi.org/10.1080/02757549608039101>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

A FUNCTIONAL APPROACH TO THE ASSESSMENT OF THE NUTRITIONAL VALUE OF PARTICULATE ORGANIC MATTER

CRISTINA MISIC^a and MAURO FABIANO^b

^a*Istituto Scienze Ambientali Marine, Università di Genova,
C. so Rainusso 14, 16038 S. Margherita L., Italy;*

^b*Cattedra di Ecologia, Università di Ancona, Via delle Brece Bianche,
60131 Ancona, Italy*

(Received 1 July 1996; in final form 30 July 1996)

Organic matter lability can be determined with different approaches, based on the quantitative and qualitative estimate of its nutritional value. The C/N ratio and the particulate organic matter (as the sum of protein, carbohydrate and lipid bulks) vs total suspended matter (POM/TSM) deal with a quantitative assessment of the organic matter bulk, while other indices such as proteins/carbohydrates ratio (PRT/CHO) may give also qualitative information. Gordon (1970) proposed a functional approach, based on organic carbon decrease after enzymatic digestion. Coupling Gordon's approach with the evaluation of the main biochemical components (proteins, carbohydrates and lipids) contained in the particulate organic matter, a simple method to assess lability and nutritional value of organic substrates is proposed.

Water samples were collected at the surface and close to the bottom in a shallow coastal station (10 m depth) in the Marconi Gulf (Ligurian Sea, NW Mediterranean). The difference between "total labile" and "hydrolyzed labile" organic matter (respectively before and after enzymatic digestion) suggests a relevant presence of resuspended refractory materials vs about 60% of labile hydrolyzed organic matter, thus confirming the importance of a more qualitative approach to organic matter studies.

Keywords: Hydrolytic enzymes; particulate organic matter; nutritional value

INTRODUCTION

The flux of particulate matter from surface productive layers of oceans is the main food source for planktonic and bottom organisms

(Fukami *et al.*, 1981; Smith *et al.*, 1992). The organic content and the nature of particles may affect the trophic pathways and the biochemical processes of the ecosystems (Amy *et al.*, 1987). Physical, chemical and biological processes modify sinking particles, enhancing or inhibiting their aggregation, influencing the solubility of organic matter and determining the settlement of chemical components (Biddanda, 1988; Biddanda and Pomeroy, 1988; Azam *et al.*, 1992). Grazing and degradation processes are the major factors responsible for depletion of energetic resources sinking to the bottom (Hobson, 1967; Cawet, 1978).

The changes in total vs labile organic matter ratio can be used as an effective nutritional value index as well as to measure detritus "aging" (Nival *et al.*, 1972; Fabiano *et al.*, 1992). Organic carbon and nitrogen assessment can offer a good nutritional value index for the organic matter, although the available fraction could be overestimated. In fact, a relevant fraction of these elements is linked to refractory compounds, like phenols or some kind of carbohydrates (Tenore and Rice, 1980; Rice, 1982). The labile fraction of organic matter may be identified with the sum of proteins, carbohydrates and lipids, measured with analytical procedures commonly employed in biochemical studies (Fabiano *et al.*, 1984; Tanoue, 1985; Fichez, 1991). Sometimes the labile fraction can be assessed based on its energetic properties (Fabiano *et al.*, 1993).

However, organic matter lability is determined by the ability of organisms to ingest and assimilate it. Assimilation processes are carried out by enzymatic activities and depend, initially, on enzyme affinity for substrates. A simple oligomeric compound is likely to resist assimilation by organisms due to poor affinity of their hydrolytic enzymes. Furthermore, a specialized organism can get energy from highly refractory substrates like vegetal debris (Stockton and DeLaca, 1982). Thus, the definition of labile substances has to take into account the hydrolysis performed by enzymes on the organic matter. Following this assumption, Gordon (1970) proposed to quantify the hydrolytic activity of some classes of enzymes (proteinases and amilases) on natural samples in order to evaluate the labile fraction in terms of carbon decrease, after digestion of the organic matter bulk.

This paper proposes a method which, matching Gordon's functional approach with the study of the changes of protein, carbohydrate and

lipid bulks, can directly measure the organic matter available to consumers. The sum of proteins, carbohydrates and lipids makes up the "total labile" organic matter, while the same components after enzymatic digestion constitute the "hydrolyzed labile" organic matter.

MATERIALS AND METHODS

Sampling

Water samples were collected at the surface and close to the bottom at a shallow coastal station (10 m depth) of the Marconi Gulf (Ligurian Sea, NW Mediterranean) with a 5 l Van Doorn bottle from November 1993 to February 1995. The samples were prefiltered through a 200 μm mesh to avoid sampling of the mesoplanktonic fraction. Sub-samples (0.5–1 litres, depending on water transparency) were then filtered through Whatman GF/F glass fibre filters (nominal pore diameter 0.45 μm), previously calcinated in a muffle furnace (450°C, 4h).

Biochemical methods

Protein analysis: The reaction suggested by Hartree (1972) for protein determination can be distinguished in two phases: copper and proteins react in a few minutes. One copper atom links up with one amino acidic residue out of four. In the second part, the Folin-Ciocalteu reactive added to the sample has a short and pH-specific reaction, because of which the sample becomes blue. Spectrophotometric analysis is undertaken as soon as possible, with 650 nm absorption light. Bovine serum albumin was used as standard. Variation coefficient: 6.6%.

Carbohydrate analysis: Carbohydrate analysis was carried out according to Dubois *et al.* (1956). The reaction between glucides and phenol in a strong acid medium (concentrated sulphuric acid) gives a colour increase proportional to glucide concentration, detectable in a spectrophotometer with 490 nm absorbance light. Standards of D(+)-glucose were utilized. The analysis coefficient of variation was 6.0%.

Lipid analysis: Lipid extraction was performed according to Bligh and Dyer (1959) with chloroform-methyl alcohol solutions. The Marsh

and Weinstein's technique (1966), based on the reaction between lipids and sulphuric acid at high temperature (200°C), was applied. The colour increase is proportional to lipid concentration. A 375 nm absorbance light was employed for spectrophotometric analysis. Tri-palmitine, dissolved in chloroform, was used as standard. Variation coefficient: 7.8%.

Carbon conversion factors: Carbon equivalents were obtained with the following conversion factors: 0.75 for lipids, 0.40 for carbohydrates and 0.49 for proteins.

Enzymatic methods

Because of the overwhelming bulk of natural hydrolytic enzymes, it is quite difficult to select three enzymatic systems capable of sustaining widespread diffusion between marine organisms and at the same time ensuring efficiency on the substrates. However, based on literature experience and recent scientific trends, trypsin (E. C. 3.4.21.4), β -glucosidase (E. C. 3.2.1.21) and lipase (E. C. 3.1.1.3), purchased from Sigma Chemicals Co. were chosen. These enzymes, since no commercial purified enzymes from marine organisms are available, are extracted from plants, bacteria and vertebrates. They have hydrolytic activities quite similar to natural marine organisms, and they are quite widespread among autotrophes and heterotrophes (Dall and Moriarty, 1983; Morton, 1983). Enzyme solutions are prepared in TRIS (hydroxymethyl aminomethane) buffer at 0.05 M. To avoid undissolved enzyme crystals, solutions were filtered through Nuclepore polycarbonate filters (0.2 μ m pore diameter). Optimal pH and temperature conditions for purified enzymes are shown in Table I. From experiments carried out to determine the best analysis conditions, proper enzyme concentration was found at 80 mg in 1000 ml TRIS. Complete hydrolysis of digestible substrates can be performed with this solution (Fig. 1). The increase in enzyme concentrations had no effect on hydrolytic processes and damaged the replicability of samples, thus increasing variation coefficients. The most effective incubation times are shown in Figure 2 (a, b and c).

Analysis procedure: Analyses were conducted with paired and one blank sample filters in their plastic petri dishes. A sample of 5 ml of

TABLE I Optimal pH, temperature and time lag of hydrolysis for trypsin, β -glucosidase and lipase

Enzyme	pH	Temp. ($^{\circ}$ C)	Time (min)
Trypsin	7.6	25	15
β -glucosidase	5.0	37	120
Lipase	7.4	37	30

enzyme solution was pipetted on to one sample filter and on to the blank one, and 5 ml of TRIS on to the other sample filter (reference sample). After hydrolysis, each filter was carefully removed from its dish, placed in a filter-holder and rinsed with the solution remaining in the dish, to return any particles that may have floated off the filter, as well as with 5 ml of deionized water. Control sample results give the initial (total) concentration of proteins, carbohydrates and lipids. Blank sample results are relatively high for proteinase assay and quite negligible for β -glucosidase and lipase. Sample results, corrected with

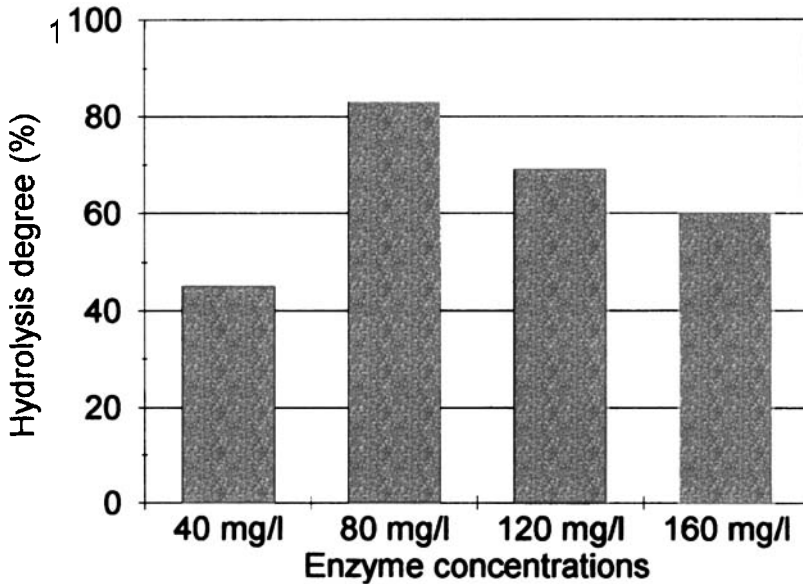


FIGURE 1 Hydrolysis (percentage) response to different trypsin concentrations in natural samples

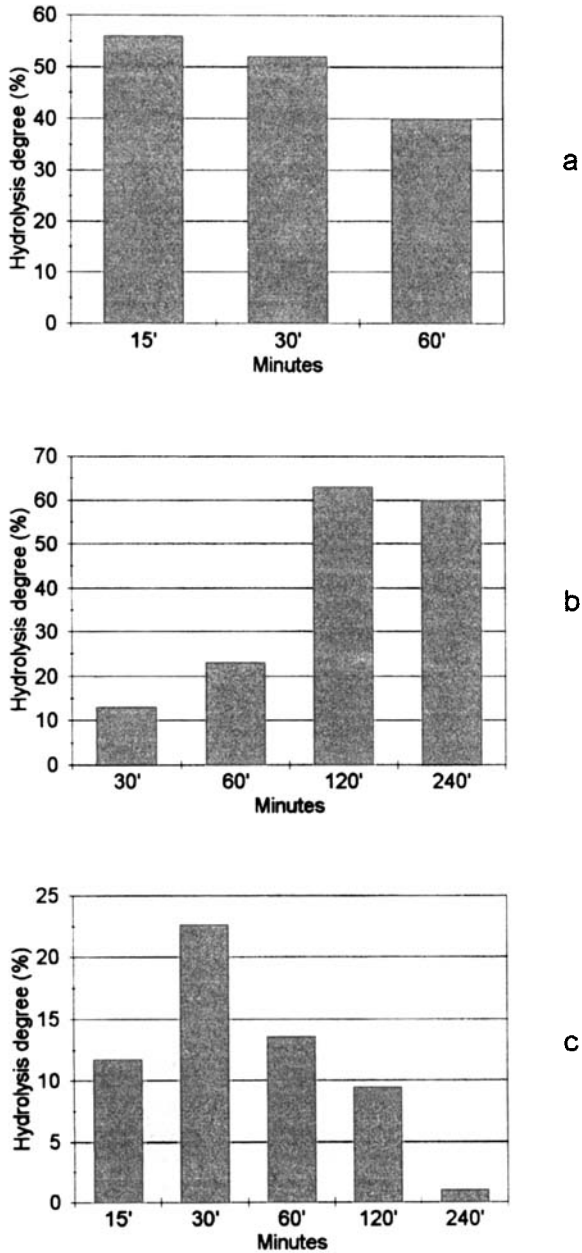


FIGURE 2 Hydrolysis response (percentage) to different incubation periods of the enzymes in natural samples. a: trypsin, b: β -glucosidase, c: lipase.

the blank results, give the concentration of refractory compounds, because of the cleavage of labile substances by enzymatic activity. The degree of hydrolysis is given by the percentage of labile vs the total compounds:

$$100 - [(SMP - BLK)/(REF - blk)] * 100$$

where:

SMP = sample concentration;

BLK = blank concentration after enzyme solution treatment

REF = control sample concentration

blk = blank concentration before enzyme solution treatment.

CONSIDERATIONS

Among the different procedures used in the literature to assess the organic matter nutritional value, the total C/N ratio has often been used for suspended particulate matter as well as for sediment organic matter (Bodungen *et al.*, 1986; Nelson and Smith, 1986; Treguer *et al.*, 1988; Fabiano *et al.*, 1993). Russell-Hunter (1970) stated that high quality organic matter is characterized by a C/N ratio of less than 17, a value based on tissue C and N contents. Thus, this could be considered as an index of appetizing food for consumers when applied to living organisms. However, it does not seem to be as effective on detrital organic matter, originating from qualitatively and quantitatively different consumption and decomposition processes.

In the Marconi Gulf waters, the C/N ratio amounts to 6.0 average in surface waters and 10.5 in the waters close to the bottom. (Tab. III). With this index, a clear distinction is made between surface and bottom waters. The significantly higher value of the examined material at the surface may relate to concern nutritional values typical of microorganisms such as bacteria (Fukami *et al.*, 1985). However, by just measuring carbon and nitrogen content, no assessment can be made on the type of molecule in which these elements are included nor can this be established if the material is labile or refractory. Therefore, the C/N ratio can be used for quantitative estimates and only marginally to understand the nutritional value of the organic matter.

TABLE II Hydrolysis percentage for proteins (PRT), carbohydrates (CHO), lipids (LIP) and particulate organic matter (POM) in superficial and bottom water

Date	Surface				Bottom			
	PRT	CHO	LIP	POM	PRT	CHO	LIP	POM
04.11.93	100.0	38.1	52.0	74.7	81.5	70.9	35.2	66.4
30.11.93	99.6	27.5	15.9	69.1	76.0	0.0	16.4	49.9
08.02.94	81.1	29.0	0.0	56.1	69.0	12.5	28.0	50.6
02.03.94	98.9	46.7	32.5	68.5	53.5	32.1	0.0	24.2
07.04.94	96.3	30.5	47.0	70.4	82.1	11.2	30.4	47.3
27.04.94	78.3	17.0	32.1	57.2	94.3	18.5	17.8	65.7
25.05.94	81.7	8.4	57.5	63.7	100.0	7.5	11.9	63.1
07.06.94	100.0	0.0	0.0	62.2	100.0	27.6	40.2	71.7
25.07.94	100.0	48.6	45.8	69.8	100.0	15.6	35.4	67.8
07.09.94	100.0	30.6	63.6	73.9	69.7	0.0	54.9	53.0
28.09.94	92.7	50.5	52.4	79.6	100.0	2.3	55.3	71.3
12.10.94	100.0	45.1	54.2	81.6	79.8	0.0	63.6	62.9
16.11.94	100.0	3.1	60.0	65.0	71.1	0.0	63.3	50.4
14.12.94	100.0	67.8	57.0	80.4	71.1	9.4	71.6	58.4
22.02.95	58.6	56.6	55.4	57.4	60.9	56.2	49.1	56.6
AVG	92.5	33.3	41.7	68.7	80.6	17.6	38.2	57.3

Similarly, the POM/TSM ratio, where POM is the sum of protein, carbohydrate and lipid bulks and TSM the total suspended matter, can be considered as a quantitative food index (Fabiano *et al.*, 1993; Navarro *et al.*, 1993). This ratio has been used successfully also on detrital organic matter. However, as far as our data are concerned, this index fails to identify any significant difference between surface and bottom water.

Referring to the POM (total)/TSM ratio (Table III), average values range between 14.5 and 16.1 for surface and bottom waters. Conversely, when considering the POM hydrolyzable portion only, these figures are

TABLE III Comparison between different food indices applied to Marconi Gulf data

Food index	Surface	Bottom
C/N	6.0	10.5
POM tot/TSM	14.5	16.1
POM hyd/TSM	9.9	9.0
PRT tot/CHO tot	2.6	3.2
PRT hyd/CHO hyd	12.5	30.3

much lower (9.9 and 9.0). This ratio, although able to give us an idea on the availability of suspended particulate matter, provides scarce information on its actual nutritional value. In fact, organic matter is used with different strategies depending on nutritional needs, and biochemical components have different features with different nutritional values for consumers.

The POM value has been used to estimate the labile fraction of particulate organic matter (Fabiano *et al.*, 1984; Tanoue, 1985; Fichez, 1991). However, although biochemical components are supposed to be digestible by organisms, the efficiency of hydrolyzing tools was not taken into account. With the hydrolytic approach, these limitations are overcome and a suitable estimate of the labile fraction as well as a good index of potential utilization of the organic matter can be obtained.

Hydrolyzed labile POM in the Marconi Gulf is about 60% of total labile matter, which provides evidence of the oligotrophic conditions in the Ligurian Sea (Table II). Hydrolysis percentage can be quite different among the three biochemical components. The protein pool is known to be more sensitive to hydrolytic attack than the other fractions, and it is likely to be utilized and modified before other more refractory products like carbohydrates, which may contain aromatic compounds (Williams and Carlucci, 1976; Hollibaugh and Azam, 1983; Newell and Field, 1983; Bodungen *et al.*, 1986; Muller *et al.*, 1986; Fabiano *et al.*, 1992). These assumptions are in agreement with the experiments carried out on enzyme hydrolytic efficiency, showing that proteinases, like trypsin, are more effective than β -glucosidase and other carbohydrate hydrolases (Antranikian, 1992). Moreover, in our experience, protein hydrolysis often reaches 100 percent (Table II).

During algae growth, the bulk of phytoplankton biomass shows a carbohydrate fraction predominance, and the newly produced proteins are quite difficult to be digested by hydrolytic enzymes because of the strong cell wall of living phytoplankton. Similarly, resuspension from the sediment of substrates linked with phenols or other compounds, which increase their refractory features, is responsible of hydrolysis degree variations. Other processes such as grazing and decomposition, modify the quantity and the chemical composition of seston particles (Fabiano *et al.*, 1992). Damaged phytoplanktonic cells, zooplanktonic faecal pellets and the bulk of excretive substances,

which characterize the post-productive periods, are a good substrate for bacterial colonization and growth (Dowgiallo, 1970; Rice and Tenore, 1981; Krog *et al.*, 1986). Moreover, hydrolysis, especially for lipids, may be related to zooplanktonic activities and to their life cycle and physiological status (Fabiano *et al.*, 1992).

The proteins/carbohydrates ratio (PRT/CHO) seems to be suitable for a qualitative study of organic matter (Fabiano *et al.*, 1993). Low values of PRT/CHO ratio, generally, indicated a mainly phytoplanktonic seston and/or the presence of an aged detritus, leading to a particulate organic matter of low nutritional quality for consumers (Nival *et al.*, 1976; Roy *et al.*, 1991; Pusceddu and Fabiano, 1996). On the other hand, the colonization of fresh algae-derived detritus by microorganisms increases the detritus nutritional value, with a higher PRT/CHO ratio (Fukami *et al.*, 1981).

The difference between total PRT/CHO ratio of surface waters in the Marconi Gulf (2.6 on average) and of bottom waters (3.2 on average) does not seem to be significant (Table III), thus leading to the same conclusions offered by a quantitative food index as POM/TSM. The hydrolyzed PRT/CHO ratio is more realistic, since it is based on the capacity of organisms to consume the organic matter and not rather its quantity. It is also a more sensitive parameter: it can detect a rather significant difference between the environment of surface (12.5) and bottom water (30.3), while stressing the importance of the area close to sediments in organic matter change processes. Despite the constant resuspension of refractory materials in this area, decomposition processes of the solid organic structures into easily assimilable materials are promoted by organic matter build-up.

CONCLUSIONS

Following the analyses carried out on samples from the Marconi Gulf, some conclusions can be drawn:

— The enzymatic method overcomes the evaluation of the labile organic matter intended as sum of proteins, lipids and carbohydrates, while it focuses on the lability and nutritional value of the main biochemical fractions of suspended matter.

— The hydrolyzed labile organic matter seems to be remarkably less abundant than labile organic matter (indicated as the sum of lipids, proteins and carbohydrates).

— The hydrolyzed PRT/CHO ratio seems to be the most suitable food index. It can give a more sensitive and realistic qualitative estimate of particulate organic matter nutritional value. This result further supports the observations made on total PRT/CHO ratio and supplies additional information to quantitative indices like POM/TSM, which failed to identify any differences between the two layers, and C/N, which gave an indication on the total amount of nitrogen containing organic material. The latter, although more abundant in surface waters, was not necessarily more appetizing.

Acknowledgements

We thank Dr. R. Danovaro and Dr. P. Povero for their suggestions and helpful comments. We are grateful to Prof. N. Della Croce who made this study possible at the Institute of Environmental Marine Sciences, University of Genova, Italy.

References

- Amy, P. S., Caldwell, B. A., Soeldner, A. H., Morita, R. Y. and Albright, L. J. (1987). Microbial activity and ultrastructure of mineral-based marine snow from Howe Sound, British Columbia. *Canadian Journal of Fisheries and Aquatic Sciences*, **44**, 1135–1142.
- Antranikian, G. (1992). Microbial degradation of starch. In: Winkelmann G. (Ed.), *Microbial Degradation of Natural Products*, **2**, 27–56.
- Azam, F., Smith, D. C. and Carlucci, A. F. (1992). Bacterial transformation and transport of organic matter in the Southern California Bight. *Progress in Oceanography*, **30**, 151–166.
- Biddanda, B. A. (1988). Microbial aggregation and degradation of phytoplankton-derived detritus in seawater. II. Microbial metabolism. *Marine Ecology Progress Series*, **42**, 89–95.
- Biddanda, B. A. and Pomeroy, L. R. (1988). Microbial aggregation and degradation of phytoplankton-derived detritus in seawater. I. Microbial succession. *Marine Ecology Progress Series*, **42**, 79–88.
- Blight, E. G. and Dyer, W. J. (1959). A rapid method for total lipid extraction and purification. *Canadian Journal of Biochemistry and Physiology*, **37**, 911–917.
- Bodungen, B., Smetacek, V., Tilzer, M. M. and Zeitzshel, B. (1986). Primary production and sedimentation during spring in the Antarctic Peninsula region. *Deep-Sea Research*, **33**, 177–194.

- Cawet, G. (1978). Organic chemistry of sea water particulates: concepts and developments. *Oceanologica Acta*, **1**, 99–105.
- Dall, W. and Moriarty, D. J. W. (1983). Functional aspects of nutrition and digestion. In: Mantel L. H. (Ed.), *The Biology of Crustacea*, Academic Press, **5**, 215–262.
- Dowgiallo, A. (1970). Water organic matter resources of high dispersion. *Polskie Archiwum Hydrobiologii*, **17**, 121–131.
- Dubois, M., Gilles, K. A., Hamilton, J. K., Rebers, P. A. and Smith, F. (1956). Colorimetric method for determination of sugars and related substances. *Analytical Chemistry*, **28**, 350–356.
- Fabiano, M., Povero, P. and Danovaro, R. (1993). Distribution and composition of particulate organic matter in the Ross Sea (Antarctica). *Polar Biology*, **13**, 525–533.
- Fabiano, M., Povero, P. and Medica, D. (1992). Carbohydrates, proteins and chlorophylls in the particulate organic matter of surface coastal waters of Ligurian Sea. *Bollettino di Oceanologia teorica ed applicata*, **10**, 41–51.
- Fabiano, M., Zavatarelli, M. and Palmero, S. (1984). Observations sur la matiere organique particulaire en Mer Ligure (chlorophylle, proteines, glucides, lipides). *Thetys*, **11**, 133–140.
- Fichez, R. (1991). Composition and fate of organic matter in submarine cave sediments; implications for the biogeochemical cycle of organic carbon. *Oceanologica Acta*, **14**, 369–377.
- Fukami, K., Simidu, U. and Taga, N. (1981). Fluctuations of the communities of heterotrophic bacteria during the decomposition process of phytoplankton. *Journal of Experimental Marine Biology and Ecology*, **55**, 171–184.
- Fukami, K., Simidu, U. and Taga, N. (1985). Microbial decomposition of phyto- and zooplankton in seawater. I. Changes in organic matter. *Marine Ecology Progress Series*, **21**, 1–5.
- Gordon, D. C. Jr. (1970). Some studies on the distribution and composition of particulate organic carbon in the North Atlantic Ocean. *Deep-Sea Research*, **17**, 233–243.
- Hartree, E. F. (1972). Determination of proteins: a modification of the Lowry method that gives a linear photometric response. *Analytical Biochemistry*, **48**, 422–427.
- Hobson, L. A. (1967). The seasonal and vertical distribution of suspended particulate matter in an area of the Northeast Pacific Ocean. *Limnology and Oceanography*, **12**, 642–649.
- Hollibaugh, J. T. and Azam, F. (1983). Microbial degradation of dissolved proteins in seawater. *Limnology and Oceanography*, **28**, 1104–1116.
- Krog, G. F., Hansen, L. and Sondergaard, M. (1986). Decomposition of lake phytoplankton. II. Composition and lability of lysis products. *Oikos*, **46**, 45–50.
- Marsh, J. B. and Weinstein, D. B. (1966). Simple charring method for determination of lipids. *Journal of Lipid Research*, **7**, 574–576.
- Morton, B. (1983). Feeding and digestion in Bivalvia. In: Saleuddin A. S. M. and Wilbur K. M. (Eds.), *The Mollusca*, Academic Press, **5**, 65–147.
- Muller, P. J., Suess, E. and Ungerer, A. (1986). Amino acids and amino sugars of surface particulate and sediment trap material from waters of the Scotia Sea. *Deep-Sea Research*, **33**, 819–838.
- Navarro, J. M., Clasing, E., Urrutia, G., Asencio, G., Stead, R. and Herrera, C. (1993). Biochemical composition and nutritive value of suspended particulate matter over a tidal flat of Southern Chile. *Estuarine, Coastal and Shelf Science*, **37**, 59–73.
- Nelson, D. M. and Smith, W. O. Jr. (1986). Phytoplankton bloom dynamics of the Western Ross Sea ice-edge. II. Mesoscale cycling of nitrogen and silicon. *Deep-Sea Research*, **33**, 1389–1412.
- Newell, R. C. and Field, J. G. (1983). The contribution of bacteria and detritus to carbon and nitrogen flow in a benthic community. *Marine Biological Letters*, **4**, 23–36.
- Nival, P., Charra, J., Malara, G. and Boucher, D. (1972). La matiere organique particulaire de la Mediterranee occidentale en Mars 1970. Mission Mediproduct du "Jean Charcot". *Annales Institut Oceanographique*, Paris, **48**, 141–156.

- Nival, P., Gostan, J., Malara, G. and Charra, J. (1976). Evolution du plancton dans la baie de Villefranche-Sur-Mer a la fin du printemps (Mai et Juin 1971). *Vie Milieu*, **26**, 47–76.
- Pusceddu, A. and Fabiano, M. (1996). Changes in the biochemical composition of *Tetraselmis suecica* and *Isochrysis galbana* during growth and decay. *Chemistry and Ecology*, **12**: 199–210.
- Rice, D. L. (1982). The detritus nitrogen problem: new observations and perspectives from organic geochemistry. *Marine Ecology Progress Series*, **9**, 153–162.
- Rice, D. L. and Tenore, K. R. (1981). Dynamics of carbon and nitrogen during the decomposition of detritus derived from estuarine macrophytes. *Estuarine, Coastal and Shelf Science*, **13**, 681–690.
- Roy, S., Mayzaud, P. and Sochu, P. (1991). Environnement physico-chimique et trophique d'un site mytilicole, Iles-de-la-Madeleine (Quebec): II-Matiere particulare, composition biochimique et productivite primaire. In: Therriault J-C (Ed.), *The Gulf of St. Lawrence: small ocean or big estuary. Canadian Special Publication of Fisheries and Aquatic Sciences*, **113**, 219–230.
- Russel Hunter, W. D. (1970). *Aquatic Productivity: an introduction to some basic aspects of biological oceanography and limnology*, Collier-MacMillan London, New-York.
- Smith, D. C., Simon, M., Alldredge, A. L. and Azam, F. (1992). Intense hydrolytic enzyme activity on marine aggregates and implications for rapid particle dissolution. *Nature*, **359**, 139–142.
- Stockton, W. L. and De Laca, T. E. (1982). Food falls in the deep sea: occurrence, quality and significance. *Deep-Sea Research*, **29**, 157–169.
- Tanoue, E. (1985). Distribution and chemical composition of POM in the Pacific sector of the Antarctic Ocean. *Trans. Tokyo University of Fisheries*, **6**, 43–57.
- Tenore, K. R. and Rice, D. L. (1980). A review of trophic factors affecting secondary production of deposit feeders. In: Tenore K. R. and Coull B. C. (Eds.) *Marine Benthic Dynamics*. University of South Carolina, 325–340.
- Treguer, P., Gueneley, S. and Kamatani, A. (1988). Biogenic silica and particulate organic matter from the Indian sector of the Southern Ocean. *Marine Chemistry*, **23**, 167–180.
- Williams, P. M. and Carlucci, A. F. (1976). Bacteria utilization of organic matter in the deep-sea. *Nature*, **262**, 810–811.